Comparison of recovered carp scales (Cyprinus carpio) gelatin and commercial calf and pork skin gelatins

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Abstract: Significant progress carries some evaluations towards the developments on seafood processing technologies and waste utilizing sector in recent years. Evaluation of wastes also has the potential to provide raw material for many industrial sectors. Evaluating the wastes also has a real potential for many industry sectors to obtain raw materials. The development of new products and commercially important of bio-molecules which have to be obtained from the wastes were important area for researches. Gelatin is used as a raw material for food industry and other industries, not only in our country but also in the world. Because of the growing demand particularly in Muslim countries hesitant considering alternatives to pork and calf sourced products are required. In the current study collagens which is used as raw material for many industries was recovered from carp scales (Cyprinus carpio). Obtained collagens was also used to produce gelatin product and some physical (colour, odour) and some functional (gelling temperature, viscosity, gel strength) properties were compared with commercial calf and pork skin gelatins.

Keywords: Gelatin, electronic noise, collagen, carp, bloom value


Anahtar kelimeler: Jelatin, elektronik burun, kollajen, sazan, bloom değeri

INTRODUCTION

Total production of gelatin was nearly 326000 metric tons in the world. 46% of the total gelatin recovered from pigskin, 29.4% from bovines, 23.1% from bones and 1.5% from other parts of the ground animals (Gómez-Guillén et al., 2002). “Gelatin” term is used for food products which were obtained from not only bones and skins of ground animals but also derived from cold blooded animals like fish by using hydrolysis techniques (Norland, 1990, Osborn et al., 1990, Grossman and Bergman, 1992, Gudmundsson and Hafsteinsson, 1997). Collagen proteins which are also known as connective tissue proteins are the source of gelatin product. By using a thermal treatment these collagen proteins can easily denaturated and converted to a gelatin (Bailey and Paul, 1998). Formed gelatin has an irreversible structure and can dissolve in water. Destruction of primary, secondary and tertiary bounds of native collagens from animals is the source of gelatin. But gelatin structure form could not be reverse again in to collagen (Fernandez-Diaz et al., 2001). Gelatin can be obtained with the partial hydrolysis of collagen which was derived from the skin, white connective tissue and also bones of animals (Morrison et al., 1999). On the other hand also can be obtained from fish skin and scale. In last decade obtained gelatins from fish became an alternative which is acceptable for halal (Muslim) and kosher (Jewish) products, these extractions have been reported previously for different fish species in the literature. The recent improvements in fish processing technology enable the converting of fish processing by-products into new value-added products or biomolecules. These motion carried the
researches to a commercially and important platform. Nearly 25% of the global fishery and processing productions are discarded as waste or processed into fish oil, fishmeal or pet food (Kim and Mendis, 2006). Most popular gelatins in the commercial sector are bovine and porcine gelatins; 60% of the market consists of these products. Due to the cultural and religious point of view consumers has some doubts and skepticism on these products. Also some part of the consumers has some health related concerns (Karim and Bhat, 2009). The utilization of aquatic resources accounts from the total was just 1% of total gelatin production (GME, 2013). And almost the origin of the gelatin obtained from the aquatic resources comes from mostly the fish intestine and fish skin (Liu et al., 2007). In 2013, the Turkish Statistical Organization (TÜİK, 2013) estimated that 8 267 tons of inland water catches come from carp (Cyprinus carpio) in Turkey. Just from this amount carp 165 tons of dry scale can be provided in Turkey. In a simple calculation this means that 49 tons dry gelatin and 705 tons of gel can be produced (6.67% w/v). The most expensive and important step in collagen recovery is removing lipid process (alcohol treatment), not only in calf and pork gelatins but also fishy odour can be a problem for consumption (Sae-leave and Benjakul, 2014). To better understand the odour effects electronic noise was used to monitor odours in the current study.

Electronic nose is a device which has a sensor array and can measure sensitively in the degree that man could not sense (Saraoglu, 2008). Electronic noise equipment can be used as a quality analyzer for gas mixtures in food industry. With using this equipment cheaper techniques can be developed when compared with other techniques (El Barbi et al., 2009). Sensors type with different characteristics such as electrochemical (metal oxide semiconductor, MOSFET), optical or piezolectric sensors (quartz crystal, surface acoustic wave) are used widely (James et al., 2005). QCM gas sensors already used in many industrial areas and also in food industry (Escuderas et al., 2011). The working style of sensors depends on frequency (frequency shift) and the proportional mass of material deposited upon the crystal (James et al., 2005). Little studies on electronic nose for gelatin odour can be seen in the literature (Muyonga et al., 2004; Ninan et al., 2014; Shyni et al., 2014). In the current study collagen was extracted from carp scales (Cyprinus carpio) and obtained collagen was used to produce gelatin product. Some physical properties like colour, odour, and some quality properties like gel strength, viscosity and gelling temperature were compared with commercial calf and pork gelatin.

**MATERIAL AND METHODS**

**Fish scale preparation**

Scales of carp (Cyprinus carpio) with an average body weight of 500 – 800 g were provided from Tan Su Ürünleri Ltd. Company in Bornova, İzmir. The scales of carp were removed by hand, one by one, samples were packaged in polyethylene bags. After filling with ice samples were quickly transported to the laboratory. Consequently, scales washed and dried by placing a table with using air condition flow. 1200 grams of scale was taken for collagen extraction and gelatin recovery.

**Gelatin extraction**

The extraction method was chosen to extract Type 1 collagens which were based on to obtain collagen proteins from the scales. Method was the combination of 3 important steps which were separation of non-collagen proteins, removing lipids from the scales and de-mineralization process. In the de-proteinization step; 5% NaCl solution (1 / 10, w/v) and 0.4% NaOH (1 / 10, w/v) were used, respectively. 1200 grams of dried scales were stirred for 30 min in room temperature with 5% NaCl solution two times. After washing scales 0.4% NaOH (1 / 10, w/v) solution was used to remove the non-collagenous proteins during 60 min. 10% Isobutyl alcohol (1 / 4, w/v) to remove lipids from the scales. This lipid removing step was repeated three times for thirty min in a digital linear shaker (Dragon Lab SK - 330 model, Beijing, China). As a final step demineralization with 0.5 N (again use %) EDTA solutions at an inherent pH 7. 66 at was performed with four different time periods; 12 h, 2 h, 2 h and 1 h (Dragon Lab SK - 330 model, Beijing, China) shaking. Between all steps, scales were collected by filtering through a sieve and washed with distilled water to separate any residual matter. Collected scales were soaked in 0.05 M acetic acid solution for 3 h. Filtered scales were placed in a tray and 1 / 3 (w/v) water was added and heated at 60 °C overnight in an oven. After filtering taken filtrate (dried thin films) was placed in a plastic tray and dried at room temperature using air condition overnight (set on 18 °C, flow temperature was determined 10 ± 2 °C). To perform the gelatin powder, dried thin films were ground using a coffee grinder. The yield of gelatin from the fish scales was calculated on a dry weight basis and expressed in %.

**Calf and pork skin gelatins**

Calf skin gelatin (Gelita GA, Germany, 1kg packs, leaves) samples were purchased from food ingredients provider company in Turkey. Because of the Turkish Laws pork products cannot be imported so same company’s commercial pork gelatin product (Gelita GA, Germany,1 kg packs, leaves) were purchased from super market chain in Hamburg, Germany.

**Proximate composition**

The moisture content (oven – drying procedure), ash content, crude protein and fat contents of samples were performed using the AOAC official methods 934. 01, 942. 01, 954. 01 and 991. 36 respectively (AOAC, 2000).

**Determination of gel strength**

GMAI (2012) standard method was used to determine the bloom strength of gelatins.

Samples were weighed and filled into the bloom bottles then dissolved inside of distilled water (55°C) to perform the final concentration of 6,67% (w/v). After keeping in the...
Comparison of recovered carp scales (Cyprinus carpio) gelatin and commercial calf and pork skin gelatins

refrigerator during 16 hours at 5°C gels were performed. Bloom strength was determined using a TAXT plus Texture analyzer (Stable Micro Systems, Godalming, UK). 25 kg load cell and 1. 27cm diameter (GL 4/P 05S) probe was used. The maximum force (in grams) recorded when the probe had penetrated 4 mm into gelatin gel’s from surface.

**Determination of viscosity**

Viscosity values of the gelatins were determined by using the method of Zhou and Regenstein (2004). Viscosity values of the gelatins were analyzed by using Brookfield DV + II Pro viscometer (Middleboro, USA). Gel solution was (6. 67% w / v) prepared at 55°C for 30 min in the magnetic stirrer until completely dissolved. HA - 4 spindle was used by helipad stand at 25°C in 60 rpm. With the help of helipad stand data were taken from vertical parts of the 100 ml gel solution.

**Gelling temperature**

Gelling temperature of the gelatins were determined by a modified (modified by Dincer et al., 2013) method of Zhou and Regenstein (2004). Brookfield DV + II Pro viscometer (Middleboro, USA) in 60 rpm with using HA - 4 spindle and helipad stand with taking the data on 60 rpm in each minute (continues test was performed until the spindle stop) was used to determine the gelling point (temperature). Data was taken between 50° C and 4°C from the 100 ml gelatin gel solution solution. Cooling was supported after reaching the room temperature with covering the sample container crashed ice. Gelling point was recorded after the spindle reached the maximum viscosity and stopped.

**Colour measurements**

Colour measurements were taken by using method of Dincer et al., (2013). Color measurement was performed by using 6,67 % (w/v) gelatin gels. Dissolved gelatin content transferred to a plastic container box and then placing in the refrigerator for16 hours at 5°C. Gel blocks were performed, procedure was used for each sample to have same smooth surface and same thickness. Preliminary measurements were taken from the surface of calibration kits to record the blind. Then gel blocks were put in to the calibration kits for measurements. Because of the transparent structure measurements were taken over standard calibration kits (Tile white and Tile black). And after calculations reference blind values were subtracted from the taken value from gel blocks.

Used calibration kit were LZM 256- Tile white (x=14.8, Y=21.2., Z=13.9) and Tile Black (x=14.5, Y=20.8, Z=15.0). By using this technique constancy was performed in measurement. Data were recorded due to the CIE Lab system, L* value which denoted lightness with a scale between 0 to 100 (black to white), a* value denoted (+) red or (−) green; and b* values denoted (+) yellow or (−) blue values were determined.

**Electronic Noise measurements**

For the E-noise measurements 2 different measurements were performed. For the first measurements gels were heated up to 55°C and then placed in to container while they were hot. And for the second measurement gels were removed from refrigerator and waited until they become in room temperature. While waiting covers of the flasks were close.

Electronic nose device used in the current study was developed by TUBITAK Marmara Research Center. Electronic nose system was the combination of; a sensor array, a A/D converter, a computer. Also pc includes software for monitoring responses (Hz) of electronic nose and for performing principal component analysis. For the sensor array, fourteen quartz crystal microbalance sensors were used due to method of Mumyakmaz et al., (2008). All gelatin samples were analyzed by two ways with the equipment. Both measurements were done in room temperature. Electronic nose measurement was performed during 60 seconds for each sample. These processes were repeated five times. The data from responses of electronic nose was processed with principal component analysis (PCA) with using software.

**Statistical analysis**

Statistical analysis were performed by using SPSS e and means comparison were done with using ANOVA test using Duncan’s multiple range tests between gelatin values.

**RESULTS AND DISCUSSION**

**Proximate composition and yield of products**

The proximate composition values of carp scales gelatin were determined as follows (dried form); 4.73 ± 0.03% moisture, 0.01 ± 0.01 % ash, 0.01 ± 0. 00% carbohydrate and 95.05 ± 1.0% protein and no fat was determined. Total yield of gelatin was calculated as 28.18% (338.16 grams of dried gelatin recovered from 1200g dry carp scales). The amount of protein in gelatin was determined respectively; 95 (carp scale), 91 (Calf skin) and 86 (Pork skin) percents in the current study. This result was found to be higher when carp scale gelatin was compared with Amur sturgeon skin gelatin 90.4% and Nile tilapia skin gelatin 88.5% (Zeng et al., 2010). Rahman et al., (2008) reported the protein content of bovine and pork gelatin as 88.7% and 90.65% respectively. But in the current commercial pork skin gelatin’s protein content was found to be lower than the mentioned study (Table 1.). As an inverse of this finding calf skin gelatin protein content was found higher with the value of 91%. The differences of the moisture contents were remarkable. Determined moisture values can be given as follows; 4.73% (carp scale), 7.13% (Calf skin) and 9.23% (pork skin) although all the gelatins form were in dried leaves. On the other hand no fat content was determined in fish scale gelatin whereas for calf and pork gelatin fat values were 1% (Table 1).

**Gel strength (Bloom value) and viscosity values**

In the current study carp scale gelatin which was produced by using acidic technique, gave us high bloom values. Taken results show that carp scale gelatin bloom value is statistically same with the calf skin gelatin bloom value and significantly higher than commercial pork gelatin (Table 2).
Table 1. Proximate chemical composition comparison of gelatins

<table>
<thead>
<tr>
<th>Samples</th>
<th>Protein (%)</th>
<th>Moisture (%)</th>
<th>Crude fat (%)</th>
<th>Crude ash (%)</th>
<th>Carbohydrate (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Carp scale gelatin</td>
<td>95.05 ± 1.0a</td>
<td>4.73 ± 0.03a</td>
<td>0.00 ± 0.00a</td>
<td>0.01 ± 0.01a</td>
<td>0.01 ± 0.00a</td>
</tr>
<tr>
<td>Calf skin gelatin</td>
<td>91.03 ± 0.2b</td>
<td>7.13 ± 0.03b</td>
<td>1.12 ± 0.12b</td>
<td>0.02 ± 0.01a</td>
<td>0.01 ± 0.00a</td>
</tr>
<tr>
<td>Pork skin gelatin</td>
<td>86.45 ± 0.4c</td>
<td>9.23 ± 0.02c</td>
<td>1.00 ± 0.05b</td>
<td>0.00 ± 0.00a</td>
<td>0.00 ± 0.01a</td>
</tr>
</tbody>
</table>

Arithmetic means and standard deviation. n = 3, different superscript letters in the same column denotes statistical difference (P<0.05)

Table 2. Gel strength (Bloom value) and viscosity comparison between gelatins

<table>
<thead>
<tr>
<th>Properties</th>
<th>Carp scale gelatin</th>
<th>Calf skin gelatin</th>
<th>Pork skin gelatin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gel strength (g)</td>
<td>341.87 ± 0.12a</td>
<td>336.87 ± 5.99a</td>
<td>308.07 ± 4.1b</td>
</tr>
<tr>
<td>Viscosity (cP)</td>
<td>26.3 ± 0.1a</td>
<td>26.7 ± 0.2a</td>
<td>6.67 ± 0.01b</td>
</tr>
</tbody>
</table>

Arithmetic means and standard deviation. GS n = 10, Vs n = 3. Different superscript letters in the same rows denotes statistical difference (P<0.05)

Bloom value (gel strength) is the most important quality criteria for gelatins. Sector and the industry prefer and expect reasonably high bloom value products (Zhou and Regenstein, 2004). In the current study gel strength of the products varied between 308 and 341 g and viscosity values varied between 26.3 and 6.67 cP (Table 2). Unexpected result was seen in pork samples; very low viscosity value was determined (Table 2) although the gel strength of the sample was over 300g that might be due to reason of some impurities. High molecular weights of non-collagen protein fractions, in the samples may decrease viscosity but not the gel strength. Also in previous studies positive correlations of gel strength and viscosity (Boran and Regenstein, 2009; Zhou and Regenstein, 2004) can be seen. Statistical analysis showed that the pork gelatin sample was significantly lower and different from other in terms of bloom value and viscosity (P<0.05). The lowest viscosity of gelatin extracted from porc skin was the lowest among the calf skin gelatin and carp scale gelatin samples. This results suggesting that carp scale can be used as an alternative raw material for gelatin production, with carrying the advantage of high viscosity and high bloom value in last gelatin product.

Gelling temperature

The gelling temperature values of the samples are given in Figure 1, Figure 2 and Figure 3 and the plots of delta (viscosity–temperature) was compared with temperature (°C) can be seen in these figures, respectively. Gelling temperature of carp scale, pork skin, and calf skin gelatin were close to each other and due to the results suitable for foods in refrigerator conditions. In figures relatively sharp increase in delta can be seen. At the summit of the delta max viscosity value can be seen in correlation with decreasing in temperature, and phase change for each sample when become 0 cp viscosity. The lowest gelling temperature was measured as 9.1 °C in pork skin gels that value was lower than the value of 10.5 °C reported for porcine skin gelatin gels by Kasankala et al., (2007). The highest gelling temperature was 13.4°C for calf skin samples, these results show that carp scale gelatin with its 10.6°C gelling point might be useful for particular food applications that require gelling temperatures like other gelatins. Supporting similar results can also be seen in the study of Boran et al., (2010).
Comparison of recovered carp scales (Cyprinus carpio) gelatin and commercial calf and pork skin gelatins

As mentioned in material section two different reference kits were used as a base in color measurements (B&W) because of the transparency of the gels. Figure 4 denotes the results of White tile values. Due to the taken results gelatins L* (Lightness) values were determined as follows, respectively; Carp scale (35.80±0.51), Calf skin (28.17±0.27) and pork skin (26.78±0.56). Due to the taken data carp scale gelatin L* value were determined significantly higher than other commercial gelatin samples. Same result can also be seen in Black tile measurement (Figure 5) of which results were respectively; 34.77±0.43, 27.56±0.70 and 27.04±0.45 for L*.

In Figures 4 and 5 as can be seen in a* parameters of color attributes no statistically difference were determined between samples (P>0.05). But in b* attributes pork skin gelatin values were significantly lower (P<0.05) than Carp scale and calf skin gelatins. Although the carp scale and calf skin were similar, a* values of carp scale showed color characteristic typical of fish gelatins, where the color tended to a little bit yellow. The L* parameter of carp scale was significantly higher than others, suggesting that carp scale gelatin was lighter than calf skin and pork skin gelatins. The difference in color among gelatins may occur due to the presence of pigment inherent in the material and depends on the raw material (Jongjareonrak et al., 2010).
Electronic Noise Values

According to the taken data due to the PCA technique (Kent et al., 2004), first principal component was determined 93% and second principal component was determined 5. Total 98% of responses from the sensors was detected. This covariance matrix of PCA was taken from multi sensor equipment showed ud 98% of the responses detected. Results of the samples according to room temperature and the 55°C responses can be seen in Figure 6. To determine the differences between the odor of the gelatin gells in 55°C, measurement was performed. Due to the taken data as shown in Figure 6. No significant difference was determined. These heated gel forms gave us very similar results. Thus explain that gelatins originated from calf skin, pork skin and carp scale smells very similar when in liquid gel forms in 55°C. As known increasing in temperature may increase the molecule speeds. Absolutely these gas molecules can easily be taken by the pumps of the E-noise but sensors could not be response easily because of the speed of odor molecules. In the literature odor comparison of the gelatins were performed by using sensorial methods instead of E-noise. In the study of Muyonga et al (2004), sensorial results did not showed a significant difference in odour between obtained Nile perch gelatin and bovine bone or commercial fish gelatins studied. Previously mentioned statement about heat and molecules may explain the reasons of no difference.

For the second measurements gells were placed in to the container while they were in room temperatures. Cover of the flask removed before placing in to the container of E-noise immediately. During the measurement both samples and the atmosphere inside of the container were equal. The responses of electronic noise measurement for calf skin, pork skin and carp scale gelatins were all determined different (Fig 6.). Similar results can also be found by Ninan et al., (2014). Although they used a sensorial method by the panelists, the odour scores were significantly higher (P < 0.05) for bovine and porcine skin gelatins than carp skin gelatins. In another study, Choi and Regenstein (2000) observed that fish gelatins had less off odor and better aroma than pork gelatins on sensory evaluation. Due to the taken results it can be concluded that both gels gave us different results depends on their origin.

**Figure 6.** E-noise measurement results of samples in 55°C and room temperature

**CONCLUSION**

In the current study obtained gelatin from carp scale was compared with two different commercial gelatin products. Taken data showed that carp scale might be successfully used as a raw material for gelatin production with an advantage of; high gel strength, viscosity, gelling temperatures, odor and color properties. Many of the functional and quality characteristics results were determined similar with calf skin and pork skin gelatins. Also advantage of the potential halal certificate may increase the marketing potential of the product. With these advantages carp scale gelatin may open to new marketing areas in Islamic and Jewish countries without any doubts of consumers. For future studies researchers should focus on decreasing the production costs. Current study designed and realized in lab conditions and many analytical degree chemicals were used. And calculated expenses showed that prices of the product were higher than commercial pork and calf skin gelatins. These expenses should decrease in future studies by using alternative chemicals, and the design of the production line should be modified for industry.

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